## Product Information

<table>
<thead>
<tr>
<th>Catalog. No.</th>
<th>Product Name</th>
<th>Format</th>
<th>Stock Conc.</th>
<th>Storage on Arrival</th>
<th>Thawing Instructions</th>
<th>Storage Once Thawed</th>
</tr>
</thead>
<tbody>
<tr>
<td>ax3755</td>
<td>Primary Human Kupffer Cells</td>
<td>1 million cells/ vial</td>
<td>N/A</td>
<td>Liquid Nitrogen</td>
<td>Follow protocol</td>
<td>N/A</td>
</tr>
<tr>
<td>ax3756</td>
<td>Kupffer Cell Plating Medium</td>
<td>250mL</td>
<td>N/A</td>
<td>4°C for 30 days</td>
<td>N/A</td>
<td>N/A</td>
</tr>
<tr>
<td>ax3757</td>
<td>Kupffer Cell Maintenance Medium</td>
<td>250mL</td>
<td>N/A</td>
<td>4°C for 30 days or 20°C for 6 months</td>
<td>Thaw at 4°C or RT</td>
<td>N/A</td>
</tr>
</tbody>
</table>

Lot-specific information such as donor information is stated in the Certificate of Analysis.

## Recommendations

- Always count the number of viable cells after thawing.
- Recommended culture vessel coating: ax3799 Type I Collagen coated multi plates.
- Recommended cell culture media: ax3756 Axol Kupffer Cell Plating Medium and ax3757 Axol Kupffer Cell Maintenance Medium.
- Recommended seeding density: 100,000 viable cells/cm².
- Recommended centrifugation speed: 500 x g for 5 min.
Primary Human Kupffer Cells

Primary Human Kupffer Cells are isolated from the whole liver tissue obtained via the gift of organ donation from donor tissue that is not suitable for organ transplantation. Human Hepatic Kupffer Cells are cryopreserved after isolation without being cultured. We do not recommend expanding or passaging these cells. Cells should be plated and used directly for endpoint assays. Do not thaw a portion of the vial to re-freeze for later use. Any other use negates the warranty.

Thawing & Plating

- Thaw the cells quickly in a 37°C water bath until just prior to complete thawing.
- Wipe the outside of the vial with 70% ethanol.
- Gently transfer the contents of the vial to a 15 mL sterile conical tube containing 8 mL of cold Plating Medium. The use of cold medium prevents cell attachment to the conical tube and minimize loss of cells during thawing process. Use of pre-warmed media is not recommended at this step.
- Rinse the cryovial with 1 mL of cold Plating Medium to ensure all the cells are transferred.
- Centrifuge the cells at 500xg for 5 min.
- Aspirate the supernatant and resuspend in 1 mL of cold Plating Medium and perform a cell count.
- Dilute the cells into the required volume of warm Plating Medium.
- Seed cells into the recommended culture vessel at the recommended seeding density and place in a humidified 37°C/5% CO₂ incubator.
- Once the cells have attached (after 4-6 h), replace the medium with warm Maintenance Medium.
- After 24 hours, replace the medium with warm Maintenance Medium and proceed with experiment.

N.B., Kupffer Cell Plating Media should not be frozen. Media is provided ready to use and prepared fresh prior to shipment. Use within 30 days of shipment.
Pathogen Testing

Samples from each donor are tested via PCR and found non-reactive to viral DNA from HIV and Hepatitis B, and viral RNA from Hepatitis C. However, since no known test can offer complete assurance, please treat the culture as a potentially infectious agent.

Product Warranty

Axol Bioscience Ltd. warrants the performance of cells only if the recommended media/reagents are used and the recommended protocols are followed. Cryopreserved cells are assured to be viable when thawed according to the recommended protocol on recommended culture ware.

Usage Statement

Our products are intended for research use only and are not to be used for any other purpose, which includes but is not limited to, unauthorized commercial uses, in vitro diagnostic uses, ex vivo or in vivo therapeutic uses or any type of consumption or application to humans.
Got any questions? Need help with the protocol? Contact Axol Technical Support at support@axolbio.com
Or
call +44 (0) 1223 751051